

PathoSEEK[®] Listeria Detection Assay with MaGiC Lysis for Detection in Cannabis Flower and MIP Matrices

Method Developer Validation

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Abstract

Background:

Current regulations require cannabis flower and cannabis products to be free of *Listeria monocytogenes*. The PathoSEEK® *Listeria* Detection Assay v2 with MaGiC Lysis Kit will detect the presence of *Listeria monocytogenes* using a qPCR (Quantitative Polymerase Chain Reaction) assay in cannabis flower, cannabis concentrates, infused edibles and infused non-edibles.

Objective:

To evaluate the PathoSEEK® *Listeria* Assay v2, using the MaGiC Lysis protocols for the presence/absence detection of *Listeria monocytogenes* in cannabis flower (delta 9-tetrahydrocannabinol >0.3%; 1g) and marijuana-infused products (MIP).

Results:

Inclusivity and exclusivity results showed the PathoSEEK® *Listeria* Detection Assay v2 method is highly specific in discriminating target organisms found in cannabis flower and infused products from non-target organisms. The MaGiC Lysis kit and PathoSEEK® *Listeria* Detection Assay v2 was successfully validated through an internal matrix study. The kit was further verified by running NSI Certified Reference Materials, part numbers CRM 7973 and FM-627. Results show that the MaGiC Lysis kit and PathoSEEK® *Listeria* Detection Assay v2 accurately detected *Listeria monocytogenes* when present.

Materials

PathoSEEK[®] Listeria Detection Assay v2 with MaGiC Lysis Kit - **P/N 420527**

(Kit contains sufficient reagents for 200 reactions)

Kit Components:

| Component Name | Qty Provided | Storage Conditions |
|--|------------------------|--------------------|
| MaGiC Lysis Reagent | 1 Bottle (12 mL) | RT |
| MaGiC Stabilization Buffer | 1 Bottle (24 mL) | RT |
| PathoSEEK [®] Amplification Mix | 3 Vials (67 rxns/each) | RT / -20 °C* |
| PathoSEEK [®] Listeria Assay v2 | 1 Tube (200 µL) | -20 °C |

Note: Actual fill volumes include overage

*The PathoSEEK Amplification Mix can be stored lyophilized at Room Temperature for up to 2 years. Once re-hydrated it must be stored at -20 °C for up to 3 months.

Additional Required Reagents Not in Kit:

| Item P/N | Item Name | Qty Provided | Storage Conditions |
|----------|--|----------------|--------------------|
| 420226 | Listeria Enrichment Broth (via Hardy Diagnostics)* | 99 mL | 2-8°C |
| 420337 | Internal Cannabis Control | 1 Tube (50 µL) | -20 °C |
| 420385 | PathoSEEK [®] Listeria Positive Control | 1 Tube (50 µL) | -20 °C |
| 420184 | PCR Grade Water | 500 mL Bottle | 2-25°C |

*Listeria Enrichment Broth is packaged in cases of 20 bottles and is drop shipped from the manufacturer. The minimum order is 4 cases. This item is a Made-To-Order product that requires a minimum order quantity as listed above. The lead time between date of order and ship date is approximately 3-4 weeks.

Method Developer Validation

Methodology

For inclusivity two different American Type Culture Collection (ATCC) *Listeria monocytogenes* species were evaluated. Strains were grown for 24 hours at $37 \pm 1^\circ\text{C}$ in *Listeria* enrichment broth. The cultures were then diluted to 100 x the Limit of Detection (LOD) of the method. For the exclusivity evaluation, a total of 45 non-targeted strains were tested. Exclusivity organisms were cultured under optimal conditions to achieve growth at the stationary phase. Inclusivity and exclusivity strains were randomized and blind coded prior to analysis.

Inclusivity and Exclusivity Evaluation

Detailed results for the inclusivity and exclusivity evaluations are presented in Tables 1 and 2. For the inclusivity evaluation, all strains of *Listeria monocytogenes* were detected by the assay using the PathoSEEK *Listeria* Detection Assay v2 with MaGiC lysis on the CFX-96, AriaMx, and the BMS Mic qPCR instruments. Table 1 shows an inclusivity study performed using an *in silico* analysis of the PathoSEEK *Listeria* Detection Assay primers against NCBI genomes. The table also includes results from testing *Listeria* species that were available from ATCC. For the exclusivity evaluation, all 45 of the organisms were correctly excluded.

Table 1: PathoSEEK[®] *Listeria* Inclusion

| Organism | Source | NCBI taxid | Max Alignment Score Percent | qPCR |
|--|---------------|-------------------|------------------------------------|-------------|
| <i>Listeria monocytogenes</i> | ATCC # 35152 | N/A | N/A | Positive |
| <i>Listeria monocytogenes</i> | ATCC # 7647 | N/A | N/A | Positive |
| <i>Listeria monocytogenes</i> FSL R2-561 | N/A | 393126 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 11-4254 | N/A | 1386005 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 88-0478 | N/A | 1196176 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2c str. 10-5026 | N/A | 1196194 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-5024 | N/A | 1196192 | 100 | N/A |
| <i>Listeria monocytogenes</i> SLCC2479 | N/A | 882020 | 100 | N/A |

| | | | | |
|--|-----|---------|-----|-----|
| <i>Listeria monocytogenes</i> serotype 1/2a str. 99-6370 | N/A | 1196179 | 100 | N/A |
| <i>Listeria monocytogenes</i> SLCC2372 | N/A | 932920 | 100 | N/A |
| <i>Listeria monocytogenes</i> N53-1 | N/A | 1234142 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 95-0093 | N/A | 1196177 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-7363 | N/A | 1386002 | 100 | N/A |
| <i>Listeria monocytogenes</i> 08-5923 | N/A | 637381 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 01-1280 | N/A | 1385996 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-7374 | N/A | 1196185 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 98-0291 | N/A | 1386007 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-6997 | N/A | 1196184 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-1046 | N/A | 1196189 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 03-5473 | N/A | 1386000 | 100 | N/A |
| <i>Listeria monocytogenes</i> 6179 | N/A | 1288295 | 100 | N/A |
| <i>Listeria monocytogenes</i> R479a | N/A | 1437838 | 100 | N/A |
| <i>Listeria monocytogenes</i> LO28 | N/A | 393132 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 02-5993 | N/A | 1196180 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 98-2035 | N/A | 1196178 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-7669 | N/A | 1196186 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 3c str. 10-5027 | N/A | 1196195 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-4758 | N/A | 1196168 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 01-6771 | N/A | 1385999 | 100 | N/A |
| <i>Listeria monocytogenes</i> SLCC 7179 | N/A | 879090 | 100 | N/A |
| <i>Listeria monocytogenes</i> Finland 1998 | N/A | 393127 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-6056 | N/A | 1196182 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-7362 | N/A | 1386001 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-6569 | N/A | 1196183 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2c str. 10-5025 | N/A | 1196193 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 3a str. 88-1059 | N/A | 1386006 | 100 | N/A |
| <i>Listeria monocytogenes</i> J0161 | N/A | 393130 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-1047 | N/A | 1196190 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-0815 | N/A | 1196188 | 100 | N/A |

| | | | | |
|--|-----|---------|-----|-----|
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-4754 | N/A | 1196167 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 01-5252 | N/A | 1385998 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-0819 | N/A | 1386004 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-0814 | N/A | 1196187 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 08-7364 | N/A | 1386003 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 04-5457 | N/A | 1196181 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 10-1321 | N/A | 1196191 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 01-1468 | N/A | 1385997 | 100 | N/A |
| <i>Listeria monocytogenes</i> 08-5578 | N/A | 653938 | 100 | N/A |
| <i>Listeria monocytogenes</i> serotype 1/2a str. 99-6871 | N/A | 1386008 | 100 | N/A |

Table 2: PathoSEEK® *Listeria* Exclusion

| Organism | ATCC Source | Origin | Results |
|-----------------------------------|-------------|---|----------|
| <i>Aeromonas hydrophila</i> | 7966 | Tin of milk with fishy odor | Negative |
| <i>Aspergillus Niger</i> | 66295 | Jerusalem artichoke tuber | Negative |
| <i>Aspergillus Tubingensis</i> | 1004 | N/A | Negative |
| <i>Bacillus amyloliquefaciens</i> | 23350 | Soil | Negative |
| <i>Bacillus subtilis</i> | 11774 | N/A | Negative |
| <i>Bacillus subtilis</i> | 23857 | N/A | Negative |
| <i>Botrytis cinerea</i> | 11542 | Azalea flowers | Negative |
| <i>Burkholderia multivorans</i> | 17616 | Soil enriched with anthranilate at 41°C | Negative |
| <i>Candida albicans</i> | 10321 | Man with bronchomycosis | Negative |
| <i>Candida glabrata</i> | 26512 | bovine placenta | Negative |
| <i>Citrobacter braakii</i> | BAA-3037 | Urine | Negative |
| <i>Citrobacter koseri</i> | 25408 | Throat | Negative |
| <i>Edwardsiella tarda</i> | 23672 | N/A | Negative |
| <i>Erwinia rhapontici</i> | 29290 | English pink wheat grains | Negative |
| <i>Escherichia hermannii</i> | 700368 | N/A | Negative |
| <i>Klebsiella oxytoca</i> | 51983 | Blood | Negative |

| | | | |
|------------------------------------|----------|---|----------|
| <i>Klebsiella pneumoniae</i> | BAA-2146 | Urine | Negative |
| <i>Lactobacillus leichmannii</i> | 4797 | N/A | Negative |
| <i>Meyerozyma guilliermondii</i> | 6260 | Sputum from patient with bronchomycosis | Negative |
| <i>Morganella morganii</i> | 25829 | Stool from Providence City Hospital | Negative |
| <i>Paecilomyces variotii</i> | 26820 | N/A | Negative |
| <i>Pantoea agglomerans</i> | 45340 | Knee laceration | Negative |
| <i>Penicillium chrysogenum</i> | 10106 | Cheese | Negative |
| <i>Penicillium citrinum</i> | 9849 | N/A | Negative |
| <i>Penicillium venetum</i> | 16025 | Hyacinthus sp. bulb | Negative |
| <i>Pichia membranifaciens</i> | 24249 | N/A | Negative |
| <i>Pityrosporum ovale</i> | 12078 | N/A | Negative |
| <i>Proteus mirabilis</i> | 43071 | Rectum | Negative |
| <i>Pseudomonas aeruginosa</i> | 35554 | N/A | Negative |
| <i>Pseudomonas aeruginosa</i> | 15442 | Water bottle in animal room | Negative |
| <i>Pseudomonas fluorescens</i> | 13525 | Pre-filter tanks | Negative |
| <i>Pseudomonas putida</i> | 47054 | N/A | Negative |
| <i>Purpureocillium lilacinum</i> | 10114 | Soil | Negative |
| <i>Ralstonia insidiosa</i> | 49129 | Clinical isolate | Negative |
| <i>Rhizopus oryzae</i> | 52748 | N/A | Negative |
| <i>Rhizopus stolonifer</i> | 2276 | N/A | Negative |
| <i>Saccharomyces cerevisiae</i> | 18824 | Top yeast | Negative |
| <i>Saccharomyces kudriavzevii</i> | 2601 | N/A | Negative |
| <i>Scopulariopsis acremonium</i> | 58636 | Chicken house soil | Negative |
| <i>Sphingomonas paucimobilis</i> | 29837 | Hospital respirator | Negative |
| <i>Staphylococcus aureus</i> | 12600 | Pleural fluid | Negative |
| <i>Staphylococcus haemolyticus</i> | 29970 | Skin | Negative |
| <i>Trichosporon mucoides</i> | 204094 | N/A | Negative |
| <i>Wickerhamomyces anomalus</i> | 2349 | N/A | Negative |
| <i>Yersinia enterocolitica</i> | 9610 | Tissue; glanders-like infection of the face | Negative |

Proficiency Testing / Certified Reference Material Results

NSI Certified Reference Materials were used to compare the detection of *Listeria monocytogenes* in the presence of different matrices. NSI part numbers used were CRM 7973 (*Listeria* Pellet) & FM-627 (*Listeria* in Hemp). Both of the pellets were enriched in *Listeria* enrichment broth for 24 hours at 37 °C. MaGiC extraction was performed in triplicate. The resulting extracted sample was then serially diluted by a factor of 10 out to 1:1,000 and qPCR was performed on non diluted sample, and each subsequent dilution. See Data in Table 3 below.

Table 3: NSI CRM Data

| Sample | Dilution | Cq Fam | Cq Hex |
|-----------------------------|----------|--------|--------|
| <i>Listeria</i> CRM | Straight | 27.59 | 26.68 |
| <i>Listeria</i> CRM | Straight | 27.59 | 26.84 |
| <i>Listeria</i> CRM | Straight | 27.86 | 26.72 |
| <i>Listeria</i> CRM | 1:10 | 27.82 | 27.02 |
| <i>Listeria</i> CRM | 1:10 | 27.45 | 26.87 |
| <i>Listeria</i> CRM | 1:10 | 27.88 | 27.04 |
| <i>Listeria</i> CRM | 1:100 | 31.61 | 27.03 |
| <i>Listeria</i> CRM | 1:100 | 31.58 | 26.91 |
| <i>Listeria</i> CRM | 1:100 | 31.32 | 27.01 |
| <i>Listeria</i> CRM | 1:1K | 34.44 | 27.15 |
| <i>Listeria</i> CRM | 1:1K | 34.58 | 27.04 |
| <i>Listeria</i> CRM | 1:1K | 35.17 | 27.02 |
| <i>Listeria</i> NSI in hemp | Straight | 28.55 | 33.57 |
| <i>Listeria</i> NSI in hemp | Straight | 28.61 | 34.01 |
| <i>Listeria</i> NSI in hemp | Straight | 28.67 | 33.34 |
| <i>Listeria</i> NSI in hemp | 1:10 | 31.25 | 35.11 |
| <i>Listeria</i> NSI in hemp | 1:10 | 31.96 | ND |
| <i>Listeria</i> NSI in hemp | 1:10 | 31.54 | 35.84 |
| <i>Listeria</i> NSI in hemp | 1:100 | 34.37 | ND |
| <i>Listeria</i> NSI in hemp | 1:100 | 34.71 | ND |

| | | | |
|-----------------------------|-------|-------|----|
| <i>Listeria NSI in hemp</i> | 1:100 | 34.28 | ND |
| <i>Listeria NSI in hemp</i> | 1:1K | 36.18 | ND |
| <i>Listeria NSI in hemp</i> | 1:1K | 36.75 | ND |
| <i>Listeria NSI in hemp</i> | 1:1K | ND | ND |
| <i>Positive Control</i> | | 12.42 | ND |
| <i>NTC</i> | | ND | ND |

Limit of Detection

An overnight culture of *Listeria monocytogenes* was diluted serially 1:10, out to 1:10 million. Next, 10ul of each dilution was plated onto 3M Petrifilm Environmental *Listeria* Plates and 10 ul was processed according to the PathoSEEK *Listeria* Assay with MaGiC Lysis protocol. The plating for the dilutions 1:10 and 1:100 were too overgrown to count and the numbers were extrapolated from the 1:1K dilution. Our MaGiC protocol was able to detect down to 2 CFUs that were visible on the 3M plates.

Table 4: LOD Results

| Sample | Dilution | MaGiC Lysis | | 3M Environmental <i>Listeria</i> Plate |
|-------------------------------|----------|-----------------|--------|--|
| | | Target Fam | Cq Fam | CFU Count |
| <i>Listeria monocytogenes</i> | 1:10 | <i>Listeria</i> | 22.72 | ~110,800 |
| <i>Listeria monocytogenes</i> | 1:100 | <i>Listeria</i> | 26.15 | ~11,080 |
| <i>Listeria monocytogenes</i> | 1:1K | <i>Listeria</i> | 29.66 | 1108 |
| <i>Listeria monocytogenes</i> | 1:10K | <i>Listeria</i> | 32.97 | 169 |
| <i>Listeria monocytogenes</i> | 1:100K | <i>Listeria</i> | 36.81 | 25 |
| <i>Listeria monocytogenes</i> | 1:1M | <i>Listeria</i> | 39.49 | 2 |
| <i>Listeria monocytogenes</i> | 1:10M | <i>Listeria</i> | ND | 1 |

***Listeria* in Marijuana-Infused Products**

An overnight culture of *Listeria monocytogenes* was spiked into 3 different MIP matrices (gummy, oil and chocolate). After 24 hours of enrichment at 37°C each sample was tested with the PathoSEEK® *Listeria* Detection Assay with MaGiC Lysis. See results in Table 5 below.

Table 5: MIP Test Results

| Sample | MIP | Target FAM | Cq FAM | Target HEX | Cq HEX |
|-------------------------------|-----------|-----------------|--------|------------|--------|
| <i>Listeria monocytogenes</i> | Gummy | <i>Listeria</i> | 21.61 | ICC | 26.59 |
| <i>Listeria monocytogenes</i> | Gummy | <i>Listeria</i> | 21.37 | ICC | 25.85 |
| <i>Listeria monocytogenes</i> | Gummy | <i>Listeria</i> | 21.33 | ICC | 26.64 |
| <i>Listeria monocytogenes</i> | Oil | <i>Listeria</i> | 20.21 | ICC | 26.58 |
| <i>Listeria monocytogenes</i> | Oil | <i>Listeria</i> | 20.50 | ICC | 25.48 |
| <i>Listeria monocytogenes</i> | Oil | <i>Listeria</i> | 20.20 | ICC | 26.22 |
| <i>Listeria monocytogenes</i> | Chocolate | <i>Listeria</i> | 21.36 | ICC | 27.06 |
| <i>Listeria monocytogenes</i> | Chocolate | <i>Listeria</i> | 21.34 | ICC | 27.16 |
| <i>Listeria monocytogenes</i> | Chocolate | <i>Listeria</i> | 21.35 | ICC | 26.72 |
| Positive Control | | <i>Listeria</i> | 13.07 | ICC | ND |
| NTC | | <i>Listeria</i> | ND | ICC | ND |

Listeria was detected in three infused cannabis matrices.

Conclusions

The PathoSEEK® *Listeria* Detection Assay with MaGiC Lysis is a rapid, alternative method to traditional plating procedures for accurate detection of *Listeria monocytogenes* in cannabis flower and cannabis infused products.

REVISION HISTORY

| Version | Date | Description |
|---------|---------------|--|
| v1 | November 2024 | Validation for PathoSEEK Listeria Detection Assay v2 |

DISCLAIMER

This test was developed, and its performance characteristics determined by Medicinal Genomics Company, for laboratory use. Any deviations from this protocol are not supported by MGC.

This test has not been validated on remediated (irradiated, ozone treated, acid treated, hydrogen peroxide treated, etc.) samples. Samples that have undergone remediation may cause discordant results between plating methods and PathoSEEK methods. When remediated samples produce a result above the action limit on qPCR, we recommend confirming viability with an approved plating method.

Results may vary based on laboratory conditions. Altitude and humidity are factors known to affect the growth of bacterial and fungal species.

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